

***Clostridium difficile* Test Algorithm: Rationale, Methods and Results**

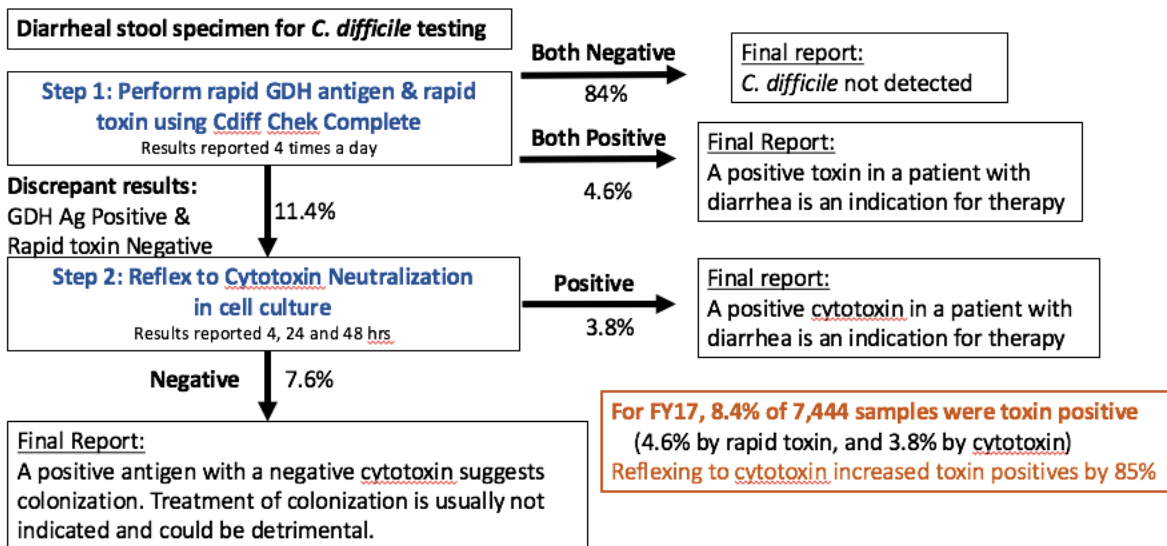
Since 2015, the *Clostridium difficile* Performance Improvement Team has spearheaded the effort to reduce *C. difficile* disease at YNHH. This has led to a dramatic 45% decrease in cases, from 9.01 to 4.95 per 10,000 patient days. Key elements of the *C. difficile* “bundle” included reducing the use of fluoroquinolones, unnecessary antibiotics, and proton pump inhibitors that drive disease, prompt and strict isolation, and stringent environmental cleaning to prevent transmission. The Laboratory assists this effort by providing prompt and accurate diagnostic test results. YNHH utilizes a unique 2-step algorithm to diagnose *C. difficile* disease, and this can lead to confusion. In this newsletter, the rationale and results for the test algorithm are presented, as well as images of the methods (**Appendix page 2**.)

Rationale: *C. difficile* infection is a toxin-mediated disease. Toxigenic strains of *C. difficile* make two toxins, A and B. Contrary to our initial understanding, Toxin B rather than toxin A is essential for disease (1,2). PCR tests determine whether a *C. difficile* strain carries the toxin B gene, but PCR does not determine whether the toxin is being actively produced *in vivo*. Several studies have shown that PCR tests over-diagnosed *C. difficile* disease and that morbidity and mortality correlated with detection of cytotoxin in stool (3-5). At YNHH, we also found that disease correlated better with cytotoxin than PCR alone (6-7). It is now recognized that toxigenic strains of *C. difficile* can colonize patients without causing disease, and that treating asymptomatic carriers can have adverse consequences (8-10).

YNHH is unusual in that **cytotoxin testing**, a diagnostic gold standard, is still available for clinical diagnosis, using cell culture plates prepared on site in the Virology Laboratory. The **cytotoxin neutralization test** is a biological assay in which the toxin damages the cells in culture, as it damages cells in the gut. To confirm specificity, the toxic effects must then be neutralized by *C. difficile* antitoxin. The downside is that cytotoxin results are read microscopically at **4, 24 and 48 hours**. In contrast, rapid toxin immunoassays are simple and fast, but lack sensitivity. The YNHH 2-step algorithm combines the speed of the immunoassays with the sensitivity of cytotoxin.

YNHH Test Algorithm: A rapid *C. difficile* GDH bacterial antigen/ toxin EIA test is performed 4 times a day in the Virology Laboratory and takes 30 minutes to complete. In FY17, **88.6%** of samples had a **final result in Step 1**: both GDH antigen and toxin negative (84%) or both positive (4.6%). If discrepant results are obtained, i.e. GDH antigen positive, but rapid toxin negative, the stool is reflexed in **Step 2** to the cytotoxin test. About 1/3 of reflexed samples are cytotoxin positive. In FY17, reflexing to cytotoxin for GDH positive samples increased the number of **toxin-positive patients** identified by 85%, from 4.6% to **8.4% of stool samples submitted**.

Algorithm at YNHH and Results from Oct 2016-Sept 2017



For FY17, 8.4% of 7,444 samples were toxin positive (4.6% by rapid toxin, and 3.8% by cytotoxin) Reflexing to cytotoxin increased toxin positives by 85%

Note: If diarrhea persists, a second stool should be submitted as toxin may be rising.

If toxin tests are negative and clinical suspicion remains high, PCR for toxin gene can be ordered by GI, ID or Infection Prevention.

C. difficile toxin gene PCR results:

No test is 100% sensitive. Therefore, Toxin gene PCR is available if antigen and/or cytotoxin tests are negative and clinical suspicion remains high. PCR can be ordered by an ID, GI or Infection Prevention attending by calling the Virology Laboratory at 688-3524.

In FY17, PCR was performed, on request, on only two patients and both were negative for *C. difficile* toxin B gene by PCR.

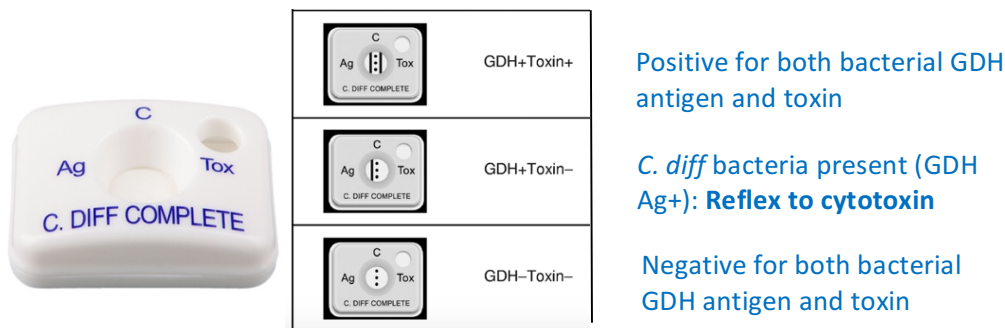
Sample submission: Liquid or semisolid stools (that conform to the contour of the container) should be submitted in leakproof containers. Solid stools will be rejected. If tests are negative, repeat testing can be ordered at 3 days. Tests for cure should not be done, as patients can remain positive despite successful therapy.

Testing is done primarily in Virology due to the expertise in cytotoxin neutralization, with Microbiology backup when Virology is closed and for *C. difficile* PCR. Routine questions should be referred to the Virology Laboratory.

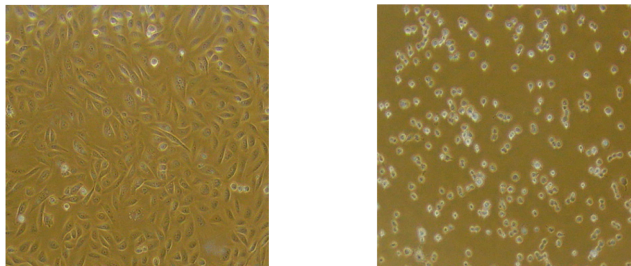
Appendix:

Test Methods:

1. **Main screening test: C. Diff Chek Complete rapid immunoassay** (done 4 times a day, ~30 minutes to result)



2. **Reflex GDH+Toxin-samples to C. diff Cytotoxin in cell culture** (biologic assay read microscopically at 4, 24 and 48 hrs)
Normal cell culture monolayer *C. difficile* cytotoxin effect on cells



3. **Xpert® C. difficile PCR:** Available on request by ID, GI, or Infection Prevention. PCR is performed in Microbiology.

Prepared by Marie L. Landry, M.D., Director Clinical Virology Laboratory.

For questions or concerns, contact the Clinical Virology Laboratory at 203-688-3524.

References

1. Cohen SH, Gerding DN, Johnson S, Kelly CP, Loo VG, McDonald LC, Pepin J, Wilcox MH, Society for Healthcare Epidemiology of A, Infectious Diseases Society of A. 2010. Clinical practice guidelines for Clostridium difficile infection in adults: 2010 update by the society for healthcare epidemiology of America (SHEA) and the infectious diseases society of America (IDSA). Infect Control Hosp Epidemiol 31:431-455.
2. Wilcox MH. 2012. Overcoming barriers to effective recognition and diagnosis of Clostridium difficile infection. Clin Microbiol Infect 18Suppl6:13-20.
3. Planche TD, Davies KA, Coen PG, Finney JM, Monahan IM, Morris KA, O'Connor L, Oakley SJ, Pope CF, Wren MW, Shetty NP, Crook DW, Wilcox MH. 2013. Differences in outcome according to Clostridium difficile testing method: a prospective multicentre diagnostic validation study of C difficile infection. Lancet Infect Dis. 13:936-945.
4. Baker I, Leeming JP, Reynolds R, Ibrahim I, Darley E. 2013. Clinical relevance of a positive molecular test in the diagnosis of Clostridium difficile infection. J Hosp Infect 84:311-315.
5. Polage CR et al. 2015. Overdiagnosis of Clostridium difficile infection in the molecular era. JAMA Internal Med 175:1792-801.
6. Kvach EJ, Ferguson D, Riska PF, Landry ML. 2010. Comparison of BD GeneOhm Cdiff real-time PCR assay with a two-step algorithm and a toxin A/B enzyme-linked immunosorbent assay for diagnosis of toxigenic Clostridium difficile infection. J Clin Microbiol 48:109-114.
7. Landry ML, Ferguson D, Topal J. Comparison of Simplexa Universal Direct PCR with Cytotoxicity Assay for Diagnosis of Clostridium difficile Infection: Performance, Cost, and Correlation with Disease. J Clin Microbiol. 52:275-80, 2014.
8. Leekha S, Aronhalt KC, Sloan LM, Patel R, Orenstein R. 2013. Asymptomatic Clostridium difficile colonization in a tertiary care hospital: admission prevalence and risk factors. Am J Infect Control 41:390-393.
9. Shim JK, Johnson S, Samore MH, Bliss DZ, Gerding DN. 1998. Primary symptomless colonisation by Clostridium difficile and decreased risk of subsequent diarrhoea. Lancet 351:633-636.
10. Kyne L, Waryn M, Qamar A, Kelly CP. 2000. Asymptomatic carriage of Clostridium difficile and serum levels of IgG antibody against toxin A. N Engl J Med 342:390-397.